

ULTRAFILTRATION PROTEIN CONCENTRATORS: CONVERSE S DEVICE

Introduction

SWISSCI Ultrafiltration Protein Concentrators have been specially designed to reduce run time and maximise protein recovery. Converse S is suitable for sample volumes of 0.5 ml up to 8 ml. Samples of 4 ml are typically concentrated within 10 minutes to 50x with macromolecular recoveries in excess of 98%. Increased concentration can be achieved with longer run time – diafiltration buffer exchange is also an option. Specially selected membranes used in Converse devices have a molecular weight cut off either 3,000, 10,000, 30,000 or 100,000 Daltons.

SWISSCI Converse S ultrafiltration devices are unique. The tubular membrane orientation and reverse direction of the flow, provide optimum cross flow conditions even for particle laden solutions. The high force moves particles and solids away from the membrane to the bottom of the device. Macromolecules collect at the base below the membrane surface; there is no risk of the filtration process allowing the sample to dry out as the system is enclosed.

No centrifuge is needed as devices can be run with specially designed Pressure Caps (to be ordered separately – see below). Results have been shown to be superior against devices run through a centrifuge, increasing the concentration speed and concentration factor.

For Pressure Cap Instructions for Use please follow Option 1. Centrifuge can be used if desired – please follow Option 2 Instructions for Use should you wish to use a centrifuge.

Option 1: Pressure Cap Operation

Converse S Concentrator use with Pressure Cap.

Equipment Required

1. Converse S Device in tube ready to use. Pipettes for sample delivery and removal.
2. Pressure Caps + 10 ml Syringe (ordered separately under product code V5-PC4).
3. Safety goggles

Option 2: Centrifugal Operation

Converse S Concentrators can be used in swing buckets or fixed angle rotors accepting standard 15 ml conical bottom tubes.

Equipment Required

1. Converse S Device in tube ready to use. Pipettes for sample delivery and removal.

Storage Conditions

Converse S should be stored in their box, at room temperature and kept out of direct sunlight.

Should you wish to reuse a device keep the membrane wet by storing with 4 ml deionised water with 1% ethanol sealed inside the Tube until you are ready to use again. Devices must be kept at room temperature and kept out of direct sunlight.

Shelf Life

Converse S has a nominal shelf life of 3 years.

OPTION 1: PRESSURE CAP INSTRUCTIONS FOR USE



Pre-Rinsing (Optional)

It is recommended to prerinse your devices with deionised water or Phosphate-Buffered Saline (PBS) solution before running through your protein samples. This will ensure the removal of Glycerine that may be present. *NOTE: Skip to Step 6 should you not require an initial pre-rinse step.*

Step 1. Remove the Red Lid from the 15 ml Tube (Fig 1) and open device.

Step 2. Place approximately 4 ml of deionised water or phosphate buffered saline into the outer section (Fig 2).

Step 3. Firmly twist on the Pressure Cap (Fig 3) and ensure the cap is on tightly to avoid any pressure loss (Fig 4). *Note: Pressure Cap must be twisted on straight.*



Fig 1.



Fig 2.



Fig 3.



Fig 4.

Step 4. Using a 10 ml syringe (Fig 5), pump the one-way valve until tube is pressurised (Fig 6-7). Leave the device for 1 minute. *Important Note: Please wear safety goggles. Once you feel resistance on the syringe the tube is pressurised. **Do not** over pressurise the device.*



Fig 5.



Fig 6.



Fig 7.

Step 5. Carefully remove the Pressure Cap from the 15 ml Tube and open device. Discard the liquid if prerinsed first.

Step 6. Pipette up to 8 ml of the sample to be concentrated into the outer section (Fig 2).

Step 7. Once the sample has been added firmly twist on the Pressure Cap (Fig 3) and ensure the cap is on tightly to avoid any pressure loss (Fig 4). *Note: Pressure Cap must be twisted on straight.*

Step 8. Repeat Steps 6-7 until all required devices have been made up.

Step 9. Using a 10 ml syringe (Fig 5), pump the one-way valve until tube is pressurised (Fig 6-7). Leave the device for approximately 10 minutes, depending on the sample solution volume and the MWCO. *Important Note: Please wear safety goggles. Once you feel resistance on the syringe the tube is pressurised. **Do not** over pressurise the device.*

Step 10. Carefully open Pressure Cap to release the pressure and remove the Cap. Remove the filtrate by placing a pipette down the centre of the device (Fig 8).



Fig 8.



Fig 9.

Step 11. Repeat Step 9-10 until the desired concentration factor has been reached and all the filtrate has been removed from each device.

Step 12. Taking one device at a time, carefully pull the Converse Device out from the Tube and recover the concentrate from the bottom of the Tube using a pipette (Fig 9) and test accordingly.

Step 13. Should you wish to reuse the device store with 4ml Deionised Water + 1% Ethanol inside the tube to keep the membrane wet.

OPTION 2: CENTRIFUGAL INSTRUCTIONS FOR USE

Pre-Rinsing (Optional)

It is recommended to prerinse your devices with deionised water or Phosphate-Buffered Saline (PBS) solution before running through your protein samples. This will ensure the removal of Glycerine that may be present. *NOTE: Skip to Step 5 should you not require an initial pre-rinse step*

Step 1. Remove the Red Lid from the 15 ml Tube (Fig 1) and open device.



Fig 1.



Fig 2.



Fig 3.

Step 2. Place approximately 4 ml of deionised water or phosphate buffered saline into the outer section (Fig 2). Carefully put the Red Lid back on tightly.

Step 3. Centrifugal spin at 2500-3500 rpm for 1 minute.

Step 4. Remove the Red Lid from the 15 ml Tube and open device. Discard the liquid if prerinsed first.

Step 5. Pipette up to 8 ml of the sample to be concentrated into the outer section (Fig 2).

Step 6. Once the sample has been added screw Red Lid back on tightly (Fig 3).

Step 7. Repeat Steps 5-6 until all required devices have been made up.

Step 8. Once sample has been added to all Tubes balance them on weighing scales to ensure the centrifuge works efficiently and without any safety issues.

Step 9. Place assembled Tubes into centrifuge and run at 2500-3500 rpm for approximately 5-15 minutes, depending on the sample solution volume and the MWCO.

Step 10. After the first spin is completed remove one device at a time from the centrifuge. Open Red Lid and remove the filtrate by placing a pipette down the centre of the device (Fig 4). Place the Red Lid back onto the device tightly once the filtrate has been removed completely.

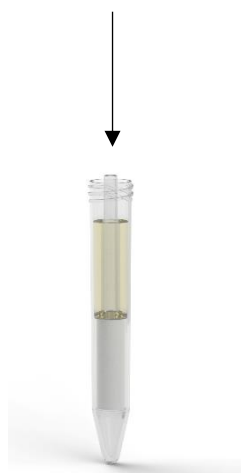


Fig 4.



Fig 5.

Step 11. Repeat Step 8-10 until the desired concentration factor has been reached and all the filtrate has been removed from each device.

Step 12. Taking one device at a time, carefully pull the Converse Device out from the Tube and recover the concentrate from the bottom of the Tube using a pipette (Fig 5) and test accordingly.

Step 13. Should you wish to reuse the device store with 4ml Deionised Water + 1% Ethanol inside the tube to keep the membrane wet.